

Fatores precipitantes na porfiria cutânea tardia no Brasil com ênfase nas mutações do gene (HFE) da hemocromatose. Estudo de 60 casos

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Abstract: BACKGROUND: Porphyria cutanea tarda is the most common form of porphyria, characterized by the decreased activity of the uroporphyrinogen decarboxylase enzyme. Several reports associated HFE gene mutations of hereditary hemochromatosis with porphyria cutanea tarda worldwide, although up to date only one study has been conducted in Brazil.

OBJECTIVES: Investigation of porphyria cutanea tarda association with C282Y and H63D mutations in the HFE gene. Identification of precipitating factors (hepatitis C, HIV, alcoholism and estrogen) and their link with HFE mutations.

METHODS: An ambispective study of 60 patients with PCT was conducted during the period from 2003 to 2012. Serological tests for hepatitis C and HIV were performed and histories of alcohol abuse and estrogen intake were investigated. HFE mutations were identified with real-time PCR.

RESULTS: Porphyria cutanea tarda predominated in males and alcohol abuse was the main precipitating factor. Estrogen intake was the sole precipitating factor present in 25% of female patients. Hepatitis C was present in 41.7%. All HIV-positive patients (15.3%) had a history of alcohol abuse. Allele frequency for HFE mutations, i.e., C282Y (p = 0.0001) and H63D (p = 0.0004), were significantly higher in porphyria cutanea tarda patients, compared to control group. HFE mutations had no association with the other precipitating factors.

CONCLUSIONS: Alcohol abuse, hepatitis C and estrogen intake are prevalent precipitating factors in our porphyria cutanea tarda population; however, hemochromatosis in itself can also contribute to the outbreak of porphyria cutanea tarda, which makes the research for HFE mutations necessary in these patients Keywords: Hemochromatosis; Hepatitis; Iron overload; Mutation; Porphyria cutanea tarda

Resumo: FUNDAMENTOS: A porfiria cutânea tardia é a forma mais comum das porfirias e caracteriza-se pela diminuição da atividade da enzima uroporfirinogênio descarboxilase. Há vários relatos da associação das mutações do gene HFE da hemocromatose hereditária com porfiria cutânea tardia no mundo, mas até hoje apenas um estudo foi realizado no Brasil.

OBJETIVOS: Estudar a associação da porfiria cutânea tardia com as mutações C282Y e H63D do gene HFE. Identificar os fatores precipitantes (hepatite C, HIV, etilismo e estrógeno) e sua relação com as mutações HFE.

MÉTODOS: Estudo ambispectivo de 60 pacientes com porfiria cutânea tardia no período de 2003 a 2012. Investigou-se as sorologias para hepatite C, anti-HIV, histórico de etilismo e ingestão de estrógenos. As mutações HFE foram identificadas com PCR em tempo real.

RESULTADOS: A porfiria cutânea tardia predominou no sexo masculino e o etilismo foi o principal fator precipitante. A ingestão de estrógenos foi o único fator precipitante em 25% das mulheres. A hepatite C estava presente em 41,7%. Todos os pacientes com HIV (15,3%) apresentavam etilismo associado. A frequência dos alelos C282Y (p=0,0001) e H63D (p=0,0004) do gene HFE foi significativamente mais elevada nos pacientes com porfiria cutânea tardia em relação à população controle. As mutações HFE não apresentavam associação com os demais fatores precipitantes.

CONCLUSÕES: Etilismo, hepatite C e ingestão de estrógenos (em mulheres) são fatores precipitantes prevalentes na nossa população com porfiria cutânea tardia, entretanto a hemocromatose isoladamente também pode contribuir para o desencadeamento da porfiria cutânea tardia, o que torna a pesquisa das mutações HFE necessária nestes pacientes.

Palavras-chave: Hemocromatose; Hepatite; Mutação; Porfiria cutânea tardia; Sobrecarga de ferro

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INTRODUCTION

Porphyria cutanea tarda (PCT) is the most common form of porphyria, characterized by decreased activity of uroporphyrinogen decarboxylase (UROD) enzyme and consequent accumulation of uroporphyrins and hepta-carboxyl porphyrinogens, predominantly in the liver. Clinically it is characterized by skin fragility, hyperpigmentation and blistering affecting photo exposed areas, facial hypertrichosis and laboratory exams indicating iron overload and increased levels of liver enzymes.² Diagnosis is based upon clinical manifestations, dosage of 24-hour urinary porphyrins (URO/COPRO > 3:1) and increased fecal levels of isocoproporphyrins (ISOCOPRO) or increase in plasma porphyrins in anuric patients.² PCT can be induced by several precipitating factors: alcoholism, estrogens, viral infections, conditions which lead to iron overload (hereditary hemochromatosis, hemodialysis, etc.) and exposure to polyhalogenated aromatic hydrocarbons triggering toxic PCT. Most patients present multiple precipitating factors. Individuals suffering from PCT are apparently genetically predisposed to developing UROD deficiency in response to liver damage.3 Iron acts as a switch that controls the generation of UROD inhibitors, commencing a vicious cycle of UROD inactivation; its removal allows restoration of UROD activity.4 Remission after phlebotomy and treatment failure when iron supplement is administered, suggests that iron has an important role in the pathogenesis of the disease.56 Interaction occurs between hereditary and acquired factors implied in UROD inactivation; however, these interactions remain undefined.7

As alcohol consumption is associated with iron overload, several studies encountered an association with HFE mutations. The pathogenic role of hepatitis C virus (HCV) in triggering PCT has been reported since 1992 and a meta-analysis of 50 studies was performed by Gisbert et al.9 The calculated average of HCV prevalence was 47% and varied depending on the country and type of PCT (57% in sporadic PCT and 26% in familial PCT).9 Association of PCT with human immunodeficiency virus (HIV) infections has been suggested; however, most of these patients have other precipitating factors, such as alcoholism, iron overload and co-infection by HCV.10 Estrogen is an independent precipitating factor for the development of PCT; no hepatic siderosis was identified in these patients. 11 There is a genetic predisposition to PCT in these patients, as 84% have decreased UROD activity, approximately 50% of normal value.³ PCT can occur in patients with chronic renal failure (CRF) undergoing hemodialysis, by preexisting decrease of hepatic UROD activity.¹²

In 1996, Feder *et al.* encountered two missense mutations, C282Y and H63D, in the HFE gene located on p21.3 region of chromosome 6p, which accounts

for most cases of hereditary hemochromatosis (HH) "type I" (88% of 178 HH patients in the study).13 Several literature studies exist which describe increased frequency of HFE gene mutations in patients with PCT (Chart 1). On chart 1 we computed the frequencies of C282Y and H63D alleles in PCT patients as well as control populations of different countries, calculating p-value. C282Y mutation originated in the population of Northwestern Europe (Celts, Vikings or Germans) and its worldwide distribution reflects migration of these populations, i.e., higher incidence occurs among Caucasians in Northwestern Europe, Canada, USA, Australia, New Zealand and South Africa.¹⁴ These mutations are rare among Asian populations, Pacific Islanders, indigenous Australians, Africans and South Americans.¹⁵ In Chile the frequency of HFE mutations in patients suffering from PCT was not different from the control group. 16 In Germany and USA the frequency of C282Y and H63D mutations is higher in patients suffering from PCT, compared to control population. 11,17,18 However, in one of the studies conducted in Germany no difference was observed in the frequency of H63D mutation.¹⁸ France and Portugal presented a greater frequency of C282Y mutation in the normal population in the North rather than in the South, probably due to the increased influence of Celtic origin in the North. 19,20 The role of S65C mutation does not appear to be decisive in PCT, as studies demonstrate that prevalence was no different between patients suffering from PCT and the control group.20 In the United Kingdom, Australia and São Paulo, Brazil, the C282Y mutation frequency was higher in PCT patients compared to controls, and no difference was observed in H63D mutation.^{21,22,23} Association between H63D mutation and PCT has been described in studies in the Mediterranean region. France, Spain, Italy, and Brazil showed a high frequency of H63D mutation in their populations. 20,23,24,25 In Spain there was a significant association of the two mutations only in sporadic PCT and not in the familial form.25

The 'major' mutation is caused by a guanine to adenine transition at nucleotide 845 of the HFE gene (845G A), which leads to replacement of cysteine by tyrosine at amino acid position 282 in the HFE protein, currently called 'C282Y' mutation. The 'minor' mutation is a replacement of cytosine by guanine at nucleotide 187 of the HFE gene (187C G), resulting in replacement of histidine by aspartic acid in amino acid position 63 of HFE protein, named 'H63D' mutation.²⁶

The C282Y mutation abolishes HFE protein interaction with transferrin receptor (TfR), which may be enough to cause an iron deposition overload.²⁷ Now mutation H63D induces a functional change,

| CHART 1: Allele frequency of C282Y and H63D mutations in control population and in porphyria cutanea tarda patients in different countries | | | | | | | | | | |
|--|--------------------|-------------|-----------------------------|------|---------|----------------|------|---|--|--|
| STUDY Author / Year | Country | Allele fre | Allele frequency in PCT (%) | | | p- value* | | | | |
| | | Nr. | C282Y | H63D | Nr. | Vr. C282Y H63D | | | | |
| Tannapfel, 2001 ¹⁷ | Germany | 115 | 1.3 | 5.2 | 190 | 30.3 | 27.9 | C282Y e H63D; p < 0.0001 | | |
| Frank, 2006 ¹⁸ | Germany | 54 | 3 | 19 | 51 | 16 | 22 | C282Y, p < 0.0013; H63D p=0.6083 | | |
| Brady, 2000 ²¹ | United Kingdom | 10566 | 8.2 | 15.3 | 84 | 36.9 | 14.9 | C282Y, p<0.0001; H63D, p=1.0000 | | |
| Stuart, 1998 ²² | Australia | Outros | | | 27 | 27.8 25.9 | | p < 0.001 for C282Y/C282Y and C282Y/H63D. | | |
| | | estudos | | | | | | Used control population of other studies. | | |
| Bulaj, 2000 ¹¹ | USA | 56 | 6.2 | 10.7 | 87 | 30.5 | 17.8 | C282Y, p<0.0001; H63D, p=0.1271 | | |
| Chiavérini, 2003 ²⁰ | Southern France | 58 | 0.9 | 12.9 | 33 | 9 | 30.3 | C282Y, p=<0.0097; H63D, p=0.0060 | | |
| Enriquez de | Spain | 50 | 2 | 18 | 69 PCTs | 10.8 | 32.6 | PCTs - C282Y p=0.0095; H63D p=0.0119 | | |
| Salamanca, 1999 ²⁵ | | | | | 19PCTf | 7.9 | 15.8 | PCTf - C282Y p=0.1281; H63D p=1.0000 | | |
| Sampietro, 1998 ²⁴ | Northern Italy | 128 | 0.7 | 12.9 | 68 | 2.2 | 29.4 | C282Y, p=0.3464; H63D, p=0.0001 | | |
| Martinelli, 2000 ²³ | São Paulo (Brazil) | 278 | 4 | 31.1 | 23 | 10.8 | 21.7 | C282Y, p=0.02; H63D, p=1.0000 | | |
| Wolff, 2006 ¹⁶ | Chile | 178 (H63D) | 1.2 | 12.4 | 20 | 7.5 20 | | C282Y, p=0.0527; H63D, p=0.2128 | | |
| | | /82 (C282Y) |) | | | | | | | |

Note: * Our calculation of p-value, considering allele frequency in each study; Nr. = Number of patients; PCTs = sporadic porphyria cutanea tarda; PCTf = familial porphyria cutanea tarda

which leads to a loss of ability to reduce affinity of TfR to transferrin. Regardless of the exact mechanism, adults with HH type 1 present a gradual increase in total body iron with increased transferrin saturation (Tsat) and serum ferritin, resulting in abnormal iron deposition, predominantly in hepatocytes but also affecting other organs. The more pronounced clinical manifestations are related to C282Y mutation and milder ones are related to H63D and S65C mutations. Absence of mutations is named wild type (WT). C282Y/WT, H63D/WT and H63D/H63D genotypes are not associated with phenotypic manifestation of the disease, although they present a small elevation of iron saturation.

Currently, the presence of homozygosis for C282Y, associated with evidence of iron overload, such as increase in concentration of hepatic iron, increase of transferrin saturation (Tsat) and ferritin levels are the gold standard for a definitive diagnosis of HH.²⁹ Gene expression (penetrance) is highly variable; not all homozygotes for C282Y will develop iron overload or HH. It is estimated that 40.0% to 70.0% of homozygous individuals for C282Y will develop some laboratory evidence of iron overload without clinical complications and that at least 50.0% of males and 25.0% of females with this genotype will develop clinical complications, secondary to iron overload.³⁰

One to 11.0% of compound heterozygotes will bring about clinical HH.31 Apparently there are gene modifiers that regulate the manifestation of HFE mutations, which when combined with individual differences of iron loss (such as females who menstruate), result in a considerable variation in clinical manifestations of risk genotypes (C282Y/C282Y and C282Y/H63D).14 In H63D homozygotes, 80.0% are clinically normal and 20.0% will develop iron overload, which is almost twice the percentage found in individuals with WT/WT genotype; H63D mutation is associated with a milder form of HH.32 The clinical symptoms of HH occur after the third decade of life, because iron deposition is gradual and cumulative. Iron build-up occurs in the liver (leading to cirrhosis and increased risk of hepatocellular carcinoma (HCC)), pancreas (diabetes mellitus), heart (heart failure and arrhythmia), osteoarticular system (arthralgia and osteoporosis) and pituitary gland (hypothyroidism, hypogonadism). Type II diabetes, joint pain and muscle fatigue are frequent manifestations.33

Most patients suffering from PCT present increased levels of serum iron and ferritin, so these changed parameters are not used to diagnose HH in these patients.^{34,35} In addition, ferritin can be increased in alcoholics and in viral infections (HCV and HIV) associated with PCT. Several authors discovered larg-

er parameters of iron levels in patients suffering from PCT, as well as C282Y homozygotes or C282Y/H63D compound heterozygotes.11 However, a German study with 190 cases of PCT demonstrated that there is no difference between serum iron, liver iron, transferrin saturation and liver enzymes in patients, with or without HFE gene mutations.17 Bulaj et al. did not observe a significant difference between the frequency of HCV in PCT patients with HFE mutations and those with wild type alleles (WT), suggesting that HFE mutations do not predispose them to HCV infection. 11 Sampietro et al. in Northern Italy observed that there was an increase in the prevalence of H63D mutation, although this mutation was not linked to the iron status.²⁴ As HCV is frequent in Italian patients suffering from PCT and rare in PCT patients of Northern Europe, Sampietro *et al.* examined a control group with active hepatitis C and without PCT, in order to discard a possible association between H63D mutation and HCV infection. Prevalence of HFE mutations in this group was identical to that observed in control groups, suggesting that the increased prevalence of H63D was in fact associated with PCT.24 The role of H63D mutation in hemochromatosis has not yet been clarified, as it is rare in classical hemochromatosis and frequent in normal individuals. Some authors suggest that H63D could be a polymorphic marker of another unknown HFE mutation.²⁴

The aims of this study cover examination of PCT association with C282Y and H63D mutations of HFE gene in Brazilian patients and identification of associated precipitating factors (alcoholism, hepatitis C, HIV infection and ingestion of estrogens), as well as their relation with presence or absence of HFE mutations.

MATERIALS E METHODS Patients

Sixty patients were included in the study, during the period from 2003 to 2011. Ethnicities considered were: Euro-Brazilian, African Brazilian, Asian Brazilian and mixed ethnicities. The comorbidities were alcoholism, estrogen intake and virus infections (HCV and HIV). We included in the clinical history exposure to chlorinated hydrocarbons and similar clinical manifestations in the family. All patients signed a free and informed consent term approved by the Committee for Research Projects Analysis of the Hospital and had blood collected for research of hemochromatosis gene mutations (C282Y and H63D). The frequency of alleles of C282Y and H63D mutations in patients with PCT was compared with frequency of alleles obtained from another study carried out in the same hospital in São Paulo (HCFMUSP), with 395 blood donors, by Pereira et al. in 2001.36

Method

PCT diagnosis was based on clinical features and quantification of urinary porphyrins (by High Performance Liquid Chromatography – HPLC) in urine sample collected for 24 hours using URO:COPRO >3:1 ratio. Liver enzymes [alanine transaminase (ALT), aspartate transaminase (AST), gamma-glutamyl transpeptidase (GGT)] and iron profile (transferrin saturation (Tsat), serum iron, ferritin, and iron binding capacity) were requested. Biochemical iron overload and liver enzyme increase were taken into consideration when values recorded were above the upper limit of normal (ULN) levels in reference values established by the Central Laboratory Division routine.

The genomic DNA leukocyte was extracted by a modified Chomczynski and Sacchi method.37 The final quantification of DNA concentration was obtained by Nanodrop™ ND-1000 Spectrophotometer reading, by 260 nm ultraviolet light absorbance and degree of DNA purity, which was evaluated and determined by ratio of 260/A280 nm. DNA whose ratio was ≥ 1.8 were used. DNA samples were stored in refrigerator at 2-8°C to be used in amplification reactions. C282Y and H63D mutation research was carried out by the real-time polymerase chain reaction (PCR-RT) technique, using 7300 Real Time PCR System ABI PRISM™ (APPLIED Ecosystem, USA). We used the detection system: Tammany® SNP Geno typing Assays (Applied Microsystems, USA), in which primers and probes labeled with fluorescent substances, in our case FAM™ and VIC®, paired in the target DNA region, allowing the identification of different alleles.

The study had an perspective and not directional outline. Data were obtained in a longitudinal form. Measures were applied in order to analyze frequency of each genotype, as well as of C282Y and H63D alleles of HFE gene of hereditary hemochromatosis (HH) in 60 patients with PCT, at 95% confidence intervals, separating the following variables (precipitating factors): alcoholism, hepatitis C virus (HCV), human immunodeficiency virus (HIV), and estrogen intake. We verified if there was an association between mutations (C282Y and H63D) and the variables listed above using Fisher's Exact Test. The adopted significance level was 5% (p<0.05). We analyzed the relation of HFE genotype with iron profile (serum iron, ferritin, transferrin saturation) and liver enzymes (AST, ALT and GGT).

RESULTS

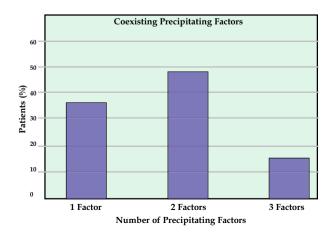
PCT predominated in males (73.3%) and there was no difference regarding the age at which the disease began in males and females (average 44.7 years).

The genotype did not interfere in age of onset of PCT. Mixed ethnicities were prevalent in most patients (72%) and the other ethnic groups were: Euro-Brazilian (23%), African Brazilian (3%) and Asian Brazilian (2%). The precipitating factors considered were: alcoholism, hepatitis C virus (HCV), estrogen intake, family history of PCT and diagnosis of hereditary hemochromatosis (HH) (considering clinical manifestation and/or presence of C282Y/C282Y or C282Y/H63D mutations). The majority of patients (63%) presented two or more coexisting precipitating factors. Graph 1 shows the distribution of number of precipitating factors coexisting in the population of 60 patients and these factors are described in table 1.

Chronic alcoholism was present in 71.7% (43 of 60 patients) and 48.8% (21 of 43) of these had HCV. Social alcoholism was referred by 10 of 60 patients (16.7%). Anti-CV was positive in 4l.7% (25 of 60 patients); 84% of these (21 of 25) were alcoholics and 20% (5 of 25) also presented positive anti-HIV. As for the HCV genotype, only nine patients were found to have it: two with genotype '1a', four with '1b' and three with '3a'. Anti-HIV was positive in 15.3% (9 of 59 patients); all patients had chronic alcoholism and five of these also presented positive anti-CV.

Only PCT family history was investigated; it was not possible to determine dosage of UROD enzyme activity in erythrocytes or search for UROD mutations in these patients. PCT case histories in the family were present in 28.3% (17 of 60 patients). Among these cases: 17.6% (3 of 17) presented this factor as the main precipitating factor; 58.8% (10 of 17) were alcoholics, 17.6% (3 of 17) hereditary hemochromatosis and 23.5% (4 of 17) estrogen intake. All cases with a PCT family history were negative for hepatitis C.

Considering 16 women in this study separately, in 50% (8 of 16) PCT was triggered by estrogens, this



GRAPH 1: Distribution of coexisting precipitating factors

being the main precipitating factor in four patients. As for the other four patients with PCT triggered by estrogen: one presented PCT family history, one had compound heterozygosity for HFE mutation (C282Y/H63D) and two had associated chronic alcoholism and family history. Still among females, in 25% (4 of 16) hepatitis C was the precipitating factor, and three of these mentioned social alcoholism. One of the females presented a previous phenotypic diagnosis of HH, which was the only precipitating factor in this patient. PCT family history was present in 31.25% (5 of 16 females), of these one featured family history as the only precipitating factor and another one featured only HFE mutation (C282Y/H63D genotype) without other precipitating factors. One female presented only social alcoholism.

The respective frequencies and confidence intervals for each precipitating factor or associated disease are described in table 2. Regarding associated diseases, two male patients were diagnosed with hepatocellular carcinoma (HCC). The first one was 60 years of age and developed HCC 32 years after onset

TABLE 1: Association of precipitating factors in 60 patients suffering from porphyria cutanea tarda

| Precipitating Factors | Nº Patients | % Patie | | |
|---------------------------------|-------------|---------|--|--|
| Alcoholism + HCV | 16 | 26.7% | | |
| Alcoholism | 9 | 15.0% | | |
| Alcoholism + FH | 7 | 11.7% | | |
| Alcoholism + HCV + HIV | 5 | 8.3% | | |
| HCV | 4 | 6.7% | | |
| Estrogens | 4 | 6.7% | | |
| Alcoholism + HIV | 3 | 5.0% | | |
| Family history of PCT (FH) | 3 | 5.0% | | |
| FH + HH | 2 | 3.3% | | |
| Alcoholism + Estrogens + FH | 2 | 3.3% | | |
| Alcoholism + HIV + FH | 1 | 1.7% | | |
| Estrogens + FH | 1 | 1.7% | | |
| Estrogens + FH + HH | 1 | 1.7% | | |
| Hereditary Hemochromatosis (HH) | 1 | 1.7% | | |
| Social Alcoholism | 1 | 1.7% | | |
| TOTAL | 60 | 100.0% | | |

Note: HCV = Positive Serology for hepatitis C virus; FH = Family History of PCT; HIV = Positive Serology for HIV virus; HH = Hereditary Hemochromatosis (C282Y/C282Y or C282Y/H63D genotypes ou HH clinical phenotype).

TABLE 2: Frequency of each precipitating factor or associated disease and its confidence interval

| Precipitating factor | NT | Estimate | CI (95%) | | | | |
|---|------|----------|-------------|-------------|--|--|--|
| or associated disease | INF. | Estimate | Lower limit | Upper limit | | | |
| Alcoholism | 43 | 71.70% | 58.60% | 82,50% | | | |
| Hepatitis C - HCV | 25 | 41.70% | 29.10% | 55,10% | | | |
| Family history | 17 | 28.33% | 17.45% | 41,44% | | | |
| HIV | 9 | 15.30% | 7.20% | 27,00% | | | |
| Diabetes mellitus | 9 | 15.00% | 7.10% | 26,60% | | | |
| Estrogen (only 9)1 | 8 | 50.00% | 24.70% | 75,30% | | | |
| Hereditary Hemochromatosis ² | 4 | 6.67% | 1.85% | 16,20% | | | |
| Hepatocellular Carcinoma | 2 | 3.33% | 0.41% | 11,53% | | | |

Note: 9: female; CI: Confidence interval; 1 Group is composed of 16 female patients; 2 Patients possessing clinical manifestation of hereditary hemochromatosis or HFE gene mutation (C282Y/C282Y or H63D/C282Y genotype).

of disease; he presented several relapses as a consequence of not stopping alcohol consumption, and alcoholism and HCV were the precipitating factors. The second patient was diagnosed with HCC at 56 years of age, 15 years after onset of disease and presented the same precipitating factors as the previously described patient. Diabetes mellitus was present in 15% (9 of 60 patients) and one of the patients had prior clinical diagnosis of hemochromatosis. The other three patients with hemochromatosis genotype (one C282Y/C282Y and two C282Y/H63D) did not present diabetes.

Research of C282Y and H63D mutations in HFE gene for HH was carried out on 60 patients. The most frequent genotype was the wild type (WT/WT), found in 25 patients (41.7%). The frequency of each genotype is described in table 3.

In our study, frequency of the C282Y allele was 8.3% and frequency of the H63D allele was 27.5%. In order to compare these findings with a control group, a study was conducted with 395 normal people who were recruited from the Blood Bank of the same Hospital (HCFMUSP), to obtain allele frequency in controls (Table 3). Allele frequency was significantly higher in PCT patients in relation to the control group for C282Y (8.3% vs. 1.77%; odds ratio 5.02; IC [95%] = [4.1%; 14.8%], p = 0.0001], p = 0.0001) and H63D (27.5% vs. 14.05%; odds ratio 2.32; IC [95%] = [19.7%; 36.4%], p= 0.0004).

Phenotypic or clinical manifestation of hemochromatosis was the only precipitating factor in the patient, who presented prior clinical diagnosis of HH despite heterozygous genotype for H63D. Two

TABLE 3: Frequency of each genotype and of C282Y and H63D alleles in 60 patients suffering from porphyria cutanea tarda and in 395 individuals of control population

| Mutations | Nr. of Patients | % Patients | | |
|-------------------------------|-----------------|------------|--|--|
| Genotype | | | | |
| C282Y / WT | 6 | 10 | | |
| C282Y / C282Y | 1 | 1.7 | | |
| H63D / WT | 21 | 35 | | |
| H63D / H63D | 5 | 8.3 | | |
| C282Y / H63D | 2 | 3.3 | | |
| WT / WT | 25 | 41.7 | | |
| Total of patients | 60 | 100 | | |
| Alleles in PCT | | | | |
| C282Y | 10/120 alleles | 8.3 | | |
| H63D | 33/120 alleles | 27.5 | | |
| Alleles in Control Population | | | | |
| C282Y | 14/790 alleles | 1.77% | | |
| H63D | 111/790 alleles | 14.05% | | |

Note: WT = Wild Type

brothers had HH diagnosis only after research of HFE mutations (a C282Y/C282Y male and a C282Y/H63D female). Another compound heterozygous patient (C282Y/H63D) is being monitored, in order to observe if clinical signs of hemochromatosis are present.

Precipitating factors were related to serology for hepatitis C and there was no association with alcoholism, estrogens, HIV, HFE mutations or HH (Table 4). The only association (p < 0.05) that occurred was that none of the cases had a family history within the HCV group.

As we analyzed if precipitating factors (HCV, HIV, alcoholism and estrogen) were associated with presence of alleles, no association was observed with C282Y or H63D alleles (Table 5). Patients with diabetes mellitus tended to have a higher frequency of H63D allele than patients without diabetes (44.4% vs. 23.5%; p-value = 0.0834; as for C282Y allele no such difference occurred. Patients with PCT family history had a tendency to an increased frequency of C282Y and H63D alleles, but it was not statistically significant (p=0.1440 for C282Y; p=0.2514 for H63D).

In table 6, as biochemical parameters of patients with and without HCV were compared, it was observed: 1. Iron levels were lower in HCV patients, but this was not statistically significant; 2. Liver enzymes were higher in HCV patients (p < 0.05).

Biochemical parameters were also distributed according to genotype in table 7. It was observed that

TABLE 4: Precipitating factors related to positive or negative serology for hepatitis C (HCV)

| Precipitating Factors | Nr. Patients tested | Patients with the precipitating factor | Patients with HCV Nr. = 25 | Patients negative for HCV Nr. = 35 | p-value |
|--------------------------|---------------------------|---|----------------------------------|---|---------|
| Alcoholism | 60 | 43 (71.7%) | 21 (84.00%) | 22 (62.85%) | 0.0886 |
| Estrogen 9 | 16 | 8 (50%) | 0 (0%) | 8 (66.67%) | 0.0769 |
| Hepatitis C - HC | V 60 | 25 (41.70%) | | | |
| HIV | 59 | 9 (15.25%) | 5 (20.83%) | 4 (11.43%) | 0.464 |
| FH | 60 | 17 (28.3%) | 0 (0%) | 17 (48.57%) | <0.0001 |
| Risk for HH ¹ | 60 | 4 (6.67%) | 0 (0%) | 4 (11.42%) | 0.1333 |
| HFE Mutations | s ² 60 | 35 (58.3%) | 13 (52%) | 22 (62.85%) | 0.4365 |
| C282Y ^{3,4} | 60 | 9 (15%) | 3 (12%) | 6 (17.14%) | 0.7222 |
| H63D ^{3,4} | 60 | 28 (46.7%) | 10 (40%) | 18 (51.42%) | 0.4386 |

Note: Nr.: number of tested patients;

♀: female;

HCV: hepatitis C virus;

HIV: human immunodeficiency virus;

FH: PCT family history;

HH: hereditary hemochromatosis.

TABLE 5: Association of precipitating factors and associated diseases with presence of C282Y and H63D alleles

| Precipitating factors / associated disease | | % with allele C282Y | p-value | % with allele H63D | p-value |
|---|---------|------------------------|---------|-----------------------|---------|
| HCV | With | 6.0% | 0.5189 | 22.0% | 0.4042 |
| | Without | 10.0% | | 30.0% | |
| HIV | With | 5.5% | >0.9999 | 11.1% | 0.1487 |
| | Without | 8.8% | | 30.0% | |
| Alcoholism | With | 6.9% | 0.4665 | 22.1% | 0.1074 |
| | Without | 11.8% | | 38.2% | |
| Estrogen | With | 6.3% | >0.9999 | 43.8% | >0.9999 |
| | Without | 6.3% | | 43.8% | |
| Diabetes | With | 5.6% | >0.9999 | 44.4% | 0.0834 |
| | Without | 8.8% | | 23.5% | |
| FH | With | 14.7% | 0.1440 | 35.3% | 0.2514 |
| | Without | 5,8% | | 23,3% | |

Table 6: Comparison between biochemical parameters (iron profile and liver enzymes) of patients with and without hepatitis C (HCV) associated to porphyria cutanea tarda

| Parameters | Nr. With | | Nr. | Without | p-value |
|---------------------|----------|-------------------|-----|-----------------|---------|
| | | HCV | | HCV | |
| Serum Iron x UNL | 18 | 0.93 ± 0.33 | 30 | 1.02 ± 0.32 | 0.463 |
| Ferritin x UNL | 18 | 1.94 ± 1.33 | 29 | 2.11 ± 1.33 | 0.702 |
| Transferrin | 17 | 46.28 ± 17.28 | 28 | 54.77 ± 17.71 | 0.058 |
| saturation (Tsat) % | | | | | |
| Serum AST x UNL | 20 | 2.57 ± 1.43 | 32 | 1.73 ± 0.82 | 0.034 |
| Serum ALT x UNL | 20 | 2.52 ± 1.46 | 32 | 1.81 ± 1.03 | 0.032 |
| Serum GGT x UNL | 20 | 4.12 ± 2.14 | 32 | 2.91 ± 2.91 | 0.015 |

Note: Nr. = Number of patients; UNL = Upper normal limit; ALT: Alanine transaminase; AST: aspartate transaminase and GGT: gamma-glutamyl transpeptidase.

TABLE 7: Iron profile (serum iron, ferritin, transferrin saturation) and liver enzymes (AST, ALT and GGT) according to HFE genotype of patient (mean ± standard deviation) in 60 patients

| Genotype | N | % | Nr | Serum Iron x UNL | Nr | Ferritin x UNL | Nr | Tsat (%) | Nr | Serum AST x UNL | Nr | Serum ALT x UNL | Nr | Serum GGT x UNL |
|-------------|----|------|----|---------------------|----|-------------------|----|-------------------|----|--------------------|----|--------------------|----|--------------------|
| C282Y/WT | 6 | 10 | 6 | 0.93 ± 0.38 | 6 | 2.25 ± 0.95 | 6 | 52.68 ± 19.41 | 6 | 2.61 ± 1.49 | 6 | 2.84 ± 1.30 | 6 | 4.55 ± 3.41 |
| C282Y/C282Y | 1 | 1.7 | 1 | 1.13 | 1 | 1.45 | 1 | 89 | 1 | 0.44 | 1 | 0.45 | 1 | 0.86 |
| H63D/WT | 21 | 35 | 17 | 1.05 ± 0.39 | 17 | 1.94 ± 1.26 | 16 | 51.48 ± 20.24 | 20 | 2.15 ± 1.24 | 20 | 2.13 ± 1.21 | 19 | 4.05 ± 3.18 |
| H63D/H63D | 5 | 8.3 | 5 | 1.12 ± 0.09 | 5 | 2.15 ± 1.21 | 5 | 63.14 ± 19.32 | 5 | 1.93 ± 0.98 | 5 | 1.61 ± 0.94 | 5 | 2.06 ± 0.56 |
| C282Y/H63D | 2 | 3.3 | 2 | 1.25 ± 0.01 | 2 | 3.17 ± 1.53 | 2 | 70.61 ± 10.76 | 2 | 1.65 ± 0.83 | 2 | 2.79 ± 1.98 | 2 | 2.29 ± 2.8 |
| WT/WT | 25 | 41,7 | 23 | 0.89 ± 0.25 | 23 | 2.10 ± 1.62 | 22 | 46.53 ± 13.28 | 25 | 1.93 ± 0.96 | 25 | 1.96 ± 1.12 | 25 | 3.60 ± 2.81 |

Note: Nr. = Number of tested patients; % of patients; UNL: upper normal limit; Tsat = transferrin saturation; UNL serum iron: $158\mu g/dL$ (male) and $145 \mu g/dL$ (female); UNL ferritin: 400 ng/dL (male) and $150\mu g/dL$ (female); normal Tsat = 20 a 40%. Due to UNL differences between males and females, we have described how many times the value was above UNL

¹ Phenotypic manifestation of patients with hemochromatosis (HH) or genotype with greater risk of HH (C282Y/H63D and C282Y/C282Y;

² Total number of patients with at least one of alleles (C282Y and H63D);

³ Present at least one C282Y and H63D mutation allele;

⁴ Compound heterozygotes entering C282Y and H63D patients.

serum iron and ferritin levels were higher in patients who were compound heterozygotes (C282Y/H63D) in relation to other genotypes. Transferrin saturation was higher in patients homozygous for C282Y (Tsat = 89%), followed by compound heterozygous (Tsat=70.61±10.76%) and homozygous for H63D (63.14 ± 19.32%). No difference was noted between genotypes in relation to liver enzymes.

DISCUSSION

Apparently an unexplained interaction between genetic and environmental factors is necessary for the PCT phenotypic expression in most cases.11 Most PCT patients presented multiple precipitating factors, of which the most common combination is between alcohol and HCV in 80% of males; in females, 40% have current estrogen intake as an isolated precipitating factor.11 In this study, the majority (63%) presented two or more coexisting precipitating factors. Chronic alcoholism was the most frequent (71.7%) precipitating factor. Alcoholism is a long-recognized important precipitating factor in PCT; it acts in synergy with other factors in predisposed individuals.38 Some authors suggest that chronic alcoholism may be associated with inheritance of HFE mutations, since C282Y mutation is dominant in countries where alcohol consumption is the most common PCT precipitating factor.35 However, in our study, no association existed between alcoholism and presence of any of the alleles; there is a greater proportion of patients with C282Y and H63D alleles among non-drinkers, but this difference did not have statistical significance.

In literature, anti-HCV antibody prevalence ranges from 8 to 91% and is related to the endemicity of HCV in the population. 1,39 In this study, positive anti-HCV was 41.7% and the majority (84%) presented association with alcoholism. Most of our patients (60%) were diagnosed for HCV infection after the emergence of PCT, hence the need to investigate HCV in these patients. Chronic alcoholism places the patient at risk of developing PCT, regardless of presence or absence of HCV. It is suggested that increasing HCV prevalence can be related to increased alcohol consumption, due to the fact that positivity for anti-HCV has been reported in up to 25% chronic drinker patients.40 However, our study has not demonstrated association of HCV with alcoholism. This study has also not observed a significant association of HCV with other precipitating factors (estrogen intake, HIV or presence of at least one HFE mutation allele). Evidently, certain individuals are predisposed to develop UROD deficiency, for the majority of individuals suffering from hepatitis C does not develop PCT.9 Liver enzymes were higher in the HCV group; however, serum iron and ferritin levels were lower in these patients. We would expect a greater increase of ferritin in patients with HCV; probably what is interfering in this finding is the fact that in the group without HCV, there are four HIV-positive patients and four patients with HFE mutations, which also contributed to increased ferritin levels.

Association with HIV occurred in nine patients (15.3%); the diagnosis was prior to PCT in six cases and simultaneous in three cases; therefore, it is always recommended to investigate HIV in these patients. As has already been mentioned in literature, it cannot be concluded that HIV alone contributes to the onset of PCT, since all patients were associated with alcoholism and five also presented HCV.⁴¹

The increased incidence in females can be attributed to estrogen intake and increased consumption of alcohol by females in recent decades. Among the 16 females, estrogens have an important role in eight (50%), and in 25% this was the only precipitating factor - similar to other studies, in which estrogen intake was the only triggering factor in more than 25% of females with PCT.³⁹ Three females had family history of PCT, which reinforces the idea that estrogen would act by inhibiting UROD, which genetically is already diminished.³

PCT family history was present in 25% of patients, but it was not possible to verify UROD erythrocyte activity or UROD gene mutations, preventing distinction between familial and sporadic PCT. This frequency is similar to that found in other studies, ranging from 20 to 29% in different populations of North America and Europe. 439 Bulaj et al. reported that inherited deficiency of UROD does not change demographic and clinical features of PCT, i.e., if the same precipitating factors (excess iron, alcohol, HCV and estrogens) contributed to clinical expression of both types of PCT (familial or sporadic), for it is unlikely that this has affected our results.³⁹

PCT is an iron-dependent disease in which iron overload is present in 60% to 70% of patients.³⁴ In PCT, transferrin saturation and ferritin are similarly increased in patients with or without HFE mutations.²² As iron always has a changed profile in patients with PCT, a research of mutations is important to identify individuals at risk for developing iron overload.³⁹

Brazilian population shows a high rate of ethnic miscegenation: the population is composed mainly of European descendants originating from Italy, Spain and Portugal, Afro-descendants (primarily from Angola, Congo and Mozambique) and native Indians; in addition, Brazil also received immigrants from Germany, Japan and the Middle East. ⁴² There are several studies reporting C282Y and H62D allele frequencies in the normal Brazilian population. ^{36,42,43} C282Y mutation prevalence among healthy individuals

(C282Y allele carrier frequency: 2.7 to 4.9%) is lower than the observed among Europeans. In relation to H63D mutation, a high prevalence exists among healthy Brazilians (frequency of allele holders: 21.6% to 39.7%); similar to that observed in White Europeans, particularly among Italians. In this study, a previous study by Pereira et al., also held at this Hospital (HCFMUSP) with 395 blood donors in 200l, was used as control group.36 Variable frequencies of C282Y and H63D alleles between different ethnic groups occur in Brazil (Euro-Brazilians, Afro-descendants and a mixture of ethnicities); frequency was high among Euro-Brazilians, which supports the theory of European origin of mutations.³⁶ As to total frequency of alleles in the normal population, we encountered 14/790 = 1.77% CI [95%] = [0.97%; 2.96%] for C282Y and 111/790 = 14,05% CI [95%] = [11.7%; 16.67%] for H63D.36 These results are very close to those found in Italian and Portuguese populations, which can be explained by the immigration observed in São Paulo.19,43,44

The prevalence of HH in Brazilian population is unknown. In Brazil, approximately one third of patients with HH classical phenotype do not present HFE mutations or other mutations associated with disease, suggesting that there may be other unknown mutations in the HFE gene or other genes involved in iron homeostasis.³⁰ These findings also match reports from Italy, where approximately one third of patients with HH do no present C282Y or H63D mutations.44 The difference in genotypes found in Brazil may be explained by Brazilian racial miscegenation, with predominant migration from Southern Europe, where mutations are less frequent, and a high influence of African descent and Native Americans, whose populations have a low frequency of these mutations.⁴³ As in Brazilian population prevalence of S65C mutation is low in the healthy population and this mutation was not found among patients with iron overload, we do not consider research of these mutations relevant in patients of this study. 30,43

Only one Brazilian study researched the association of HFE mutation (C282Y and H63D) in PCT. ²³ In 23 PCT patients, four patients presented C282Y mutation (27.4%) and seven patients presented H63D mutation (30.4%). The majority was heterozygous for the mutation; only one was homozygous for H63D mutation; one was compound heterozygous (C282Y/H63D) and none homozygous for C282Y. In the control group, 278 individuals were observed: C282Y mutation was found in 4% and H63D mutation in 31.1%. Comparing both groups, there was a significant statistical difference between the two groups for C282Y mutation (OR=5.1, 95% IC 1.5-17.6, p = 0.02), but it was not the case for H63D (OR=1, 95% IC 0.4-

2.4, p = 1).²³ In our study, the frequency of two mutations was significantly higher in PCT patients in relation to control population for C282Y (p=0.0001) and for H63D (p= 0.0004). We may conclude that HFE mutations increase the susceptibility of individuals to PCT; iron buildup precipitates manifestation of PCT in individuals already predisposed to the disease. The association of iron and chronic hepatitis C led to investigation of HFE mutations as a contributing factor for progression of liver disease, but the results are conflicting. Some researches report the association of HFE mutations with severe liver disease, while others have not found this association.8 The HCV infection and the C282Y mutation are independent precipitating factors, for in a study of Stuart et al. all HCV patients were homozygous for the wild type²² and in a study of Toll et al. C282Y prevalence was higher among patients without HCV.45 In our study, no association was observed between precipitating factors (HCV, HIV, alcoholism and estrogen) and presence of C282Y and H63D alleles. Patients with a PCT family history tended to have an increased frequency of C282Y and H53D alleles, but it was not statistically significant (p=0.1440 and p=0.2514, respectively); allele inheritance may contribute in part with PCT family history. There has been a tendency to an increase of H63D allele among patients with diabetes mellitus (p=0.0834), but not for C282Y alleles. This finding may be due to the fact of high prevalence of H63D mutation, present in the normal Brazilian population.

HH presents few specific signs or symptoms to alert the physician. The symptoms are nonspecific (e.g. arthralgia, weakness and fatigue) and do not help when there is suspicion of HH. 46 Symptoms usually develop in patients over 40 years of age, at a time when iron deposition in organs may have caused irreversible damage in tissues, such as cirrhosis, hepatocellular carcinoma, arthritis, hypogonadism, diabetes mellitus and cardiomyopathy.47 One of the patients in this study had prior clinical diagnosis of HH as single precipitating factor of PCT. This patient was Oriental and heterozygous for H63D; probably other unknown mutations related to HH were contributing to the outbreak of PCT in this case. Neither the homozygous C282Y patient nor his compound heterozygous sister presented any other precipitating factors, except HFE mutations and family history. These three patients indicate that HFE mutations or clinical manifestation of hemochromatosis may contribute to the triggering of PCT, regardless of the presence of other precipitating factors.

The phenotypic expression of HH is higher in homozygous patients for C282Y and in compound heterozygotes. ¹¹ Through discovery of the HFE gene, it

was possible to identify 'potential' risk of developing iron overload, for not all C282Y homozygotes will develop iron overload or HH, and it is not possible to predict the degree of phenotypic expression. In our study, it was observed that serum iron and ferritin levels were higher in compound heterozygous patients (C282Y/H63D) in comparison to other genotypes, and there was no difference in liver enzymes between genotypes. As for transferrin saturation, it was higher in homozygous C282Y patients (Tsat = 89%), followed by compound heterozygous (Tsat = 70.61±10.76%) and H63D homozygotes (Tsat = 63.14%), indicating that this parameter may be important when suspicion of association with HFE mutations arises regarding PCT patients. HH is suspected when transferrin saturation is >50% in premenopausal females and >60% in males or postmenopausal females but the elevation of Tsat is not specific to HH.48 Elevation of Tsat is the first manifestation of HH and may be high in young individuals, who still possess normal ferritin.49 In order to quantify iron overload in HH, we usually use plasma ferritin, but this interpretation is distorted in PCT and in several other infectious and inflammatory diseases (acute phase reactant), e.g. in alcoholic hepatitis, nonalcoholic steatohepatitis, hepatitis C, rheumatoid arthritis and neoplastic diseases such as lymphoma.49 In patients with HFE mutations or clinical manifestation of HH, which present factors that interfere with plasma ferritin, we usually suggest use of magnetic resonance imaging (MRI) of the liver in order to quantify liver iron concentration, a noninvasive method with a high degree of sensitivity and preciseness in order to measure iron concentration, as well as providing information about degree of fibrosis.⁵⁰ One should always monitor iron levels in PCT patients (regardless of presence of HFE mutation), even when they are in a biochemical remission of disease, for these patients may present unknown factors that lead to an increase in iron level and subsequently lead to PCT recurrence.

CONCLUSION

Alcoholism, hepatitis C and estrogen intake (females) are precipitating factors, prevalent in PCT Brazilian population, but HH alone may also contribute to the onset of PCT, which makes research of HFE mutations or HH phenotypes in these patients important .

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